Group A Beta Strep Screen
Depress the tongue and pass the swab firmly over the back of the patient's throat, tonsils or tonsillar fossae and any area of inflammation and or exudation. Continue with Step 2 below

Wound Specimens
1. Pass the swab firmly over or into an area of suspected infection and obtain a sample of exudate, drainage, or purulent discharge if these are present.
2. Return the swab to the transport tube and break the media ampule at the base of the tube to moisten the swab.
3. Label the swab transport tube with patient name, date and specimen source. Indicate the area of the body from which the wound was taken to assist in distinguishing normal flora (ear, leg, etc)

Stool Culture for Campylobacter, Salmonella, Shigella, and Shigatoxin E.coli. (Para-Pak C&S)
If other pathogens are suspected please indicate on request form.
1. Obtain a collection kit containing Cary Blair or stool transport media.
2. Specimens may be obtained by covering the rear half of the toilet rim below the seat with plastic kitchen wrap to catch feces.
3. Transfer feces (tablespoon size portion) into the kit up to the line and mix thoroughly. Label container with patient name and date. Store samples in refrigerator

Norovirus cannot be tested by stool culture.
For Norovirus stool PCR- collect feces in clean leak-proof container with no preservatives. Label container with patient name and date.
Store samples in refrigerator

Urine Culture Collection
1. Wash hands with soap and water, rinse and dry.
2. WASH area around urethra with soap.
3. RINSE area with warm water.
4. VOID- Pass the first portion of urine into the toilet and then pass a portion (1 ounce) of the remaining urine into a sterile container. Pass the rest of the urine into the toilet, close and label the container with name and date. Store samples in refrigerator

Complete the requisition form or order test directly in LIMS. Place the specimen and requisition form in the transport pouch and arrange for courier pickup. Specimens collected should be delivered to laboratory within 24 hours after collection for best results. Stool and Culturettes that are 24-72 hours old are acceptable if refrigerated. Courier specimen pick up locations can be found at the following web address: http://www.dhss.delaware.gov/dhss/dph/lab/courierroutes.html. Contact the Lab for specific collection of other sources at 302-223-1520 Revised 32817